

# Novel Production Strategies for Biosurfactants

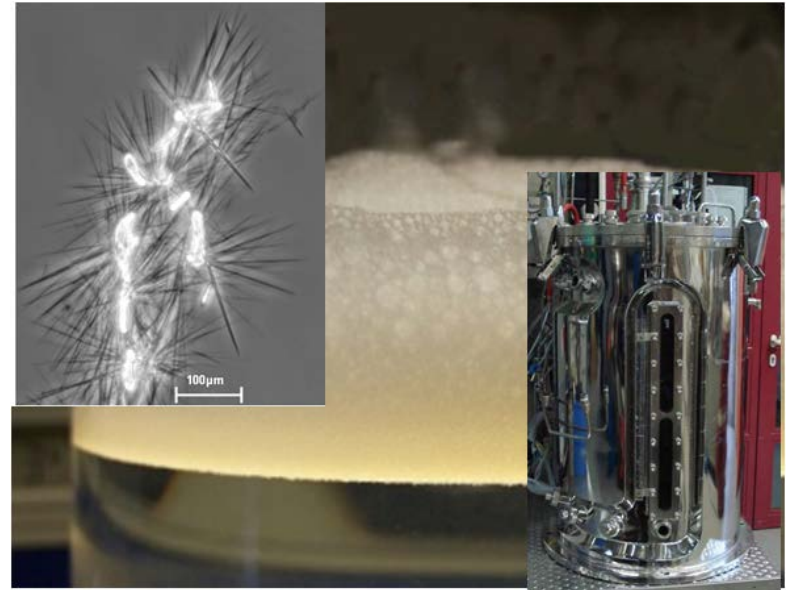
Project acronym: BioSurf.  
Project no: EIB.10.039



## Project aim

Surfactants form an integral part of our everyday life with applications from food to fuel additives

BioSurf aims at an increased replacement of petro-based surfactants by biosurfactants generated from renewable resources





# Overview of the WPs

BioSurf: Novel Production Strategies for Biosurfactants



## WP2

Identification and preparation of new surfactants



## WP 3

Enzymatic synthesis and modification



## WP 4

Product characterization and selection



## WP 5

Metabolic engineering  
Improved strains for application



## WP 6

Bioprocess engineering and downstream processing



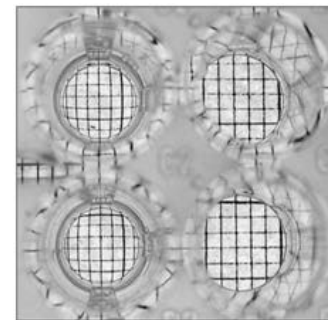
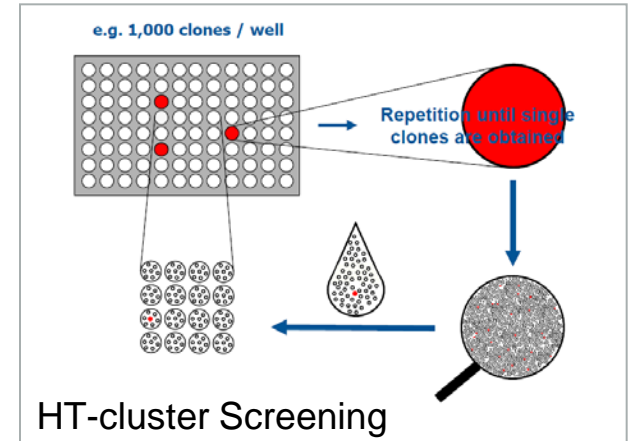
## WP 7

Dissemination and Exploitation



# WP2: Identification and preparation of new surfactants

- Development of modified or unexplored biosurfactants
- Identification of novel biosurfactants by screening of strain collections
- Characterisation of microorganisms and the produced biosurfactants



Microplate assay based on surface activity of BS

# WP2: Identification and preparation of new surfactants



## 1. Screening for new surfactant producers:

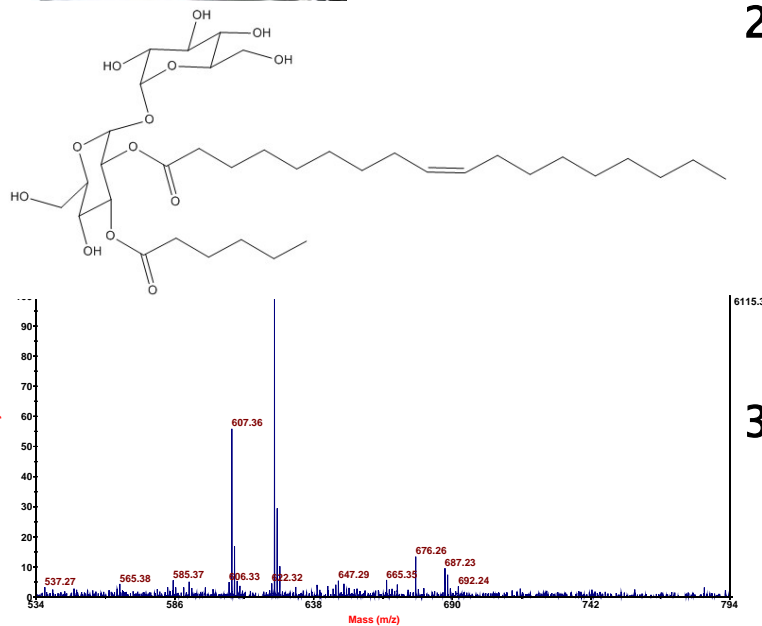
- Establishment of high throughput screening methods
- Isolation and screening of 377 microorganisms (e.g. peat-bog soil, intestines of seawater animals, mediterranean sponges)
- Identification of not yet described non-pathogenic biosurfactant producers (*Rahnella sp.*, *Citrobacter braaki*, *Serratia fonticola*, *Pseudomonas extremaustralis* and several *Psychrobacter sp.*).

## 2. Production and purification

- Fermentation (L scale) of novel Trehalose-lipids from *Tsukamurella spumae* and *T. pseudospumae*
- Production of unknown surfactants (584 and 600Da) from *Rahnella sp.* 323 (1,2 L scale)

## 3. Characterization and structure elucidation

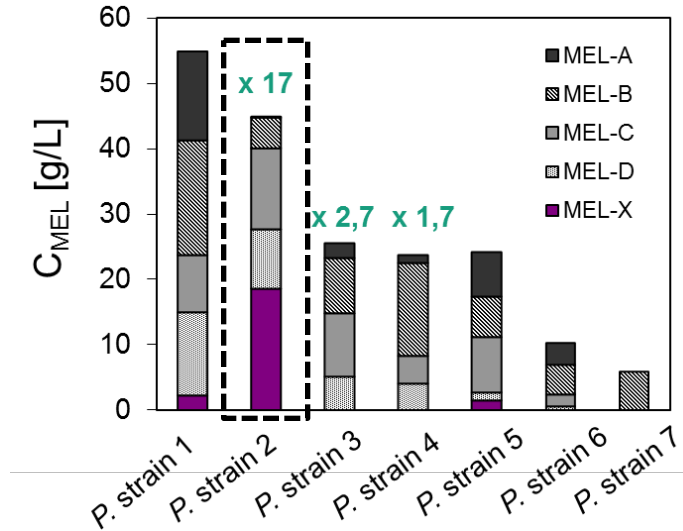
- Isolation by liquid chromatography  
structure elucidation using MALDI-ToF-MS-MS and NMR



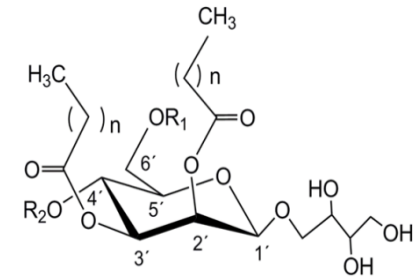


# WP2: Production of rare hydrophilic MEL-structures

- Comparison of 7 *Pseudozyma* strains
- ➔ 5 produce abundant amounts of MEL-B, MEL-C MEL-D and with MEL-X greater hydrophilicity

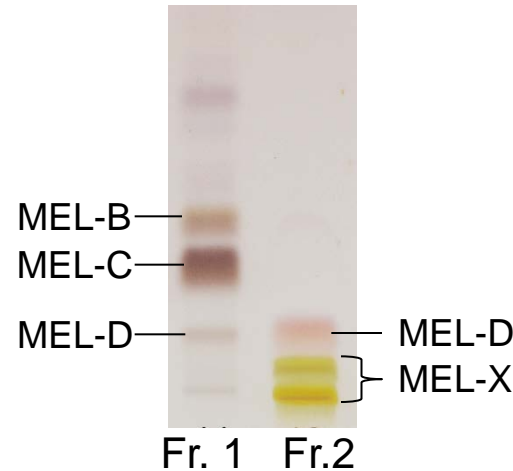


n = 5-16  
 MEL-A: R<sub>1</sub>, R<sub>2</sub> = Acetyl  
 MEL-B: R<sub>1</sub> = Acetyl, R<sub>2</sub> = H  
 MEL-C: R<sub>1</sub> = H, R<sub>2</sub> = Acetyl  
 MEL-D: R<sub>1</sub>, R<sub>2</sub> = H



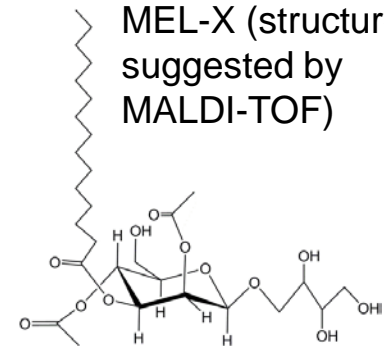
- *P. strain 2* produces a 17-fold higher MEL-yield, compared to literature (2-fractions)

1. MEL-B, -C, -D-fraction (20 ± 5 g/L)
2. MEL-D, -X-fraction with high water solubility (25 ± 5 g/L)



monoacylated

MEL-X (structure suggested by MALDI-TOF)

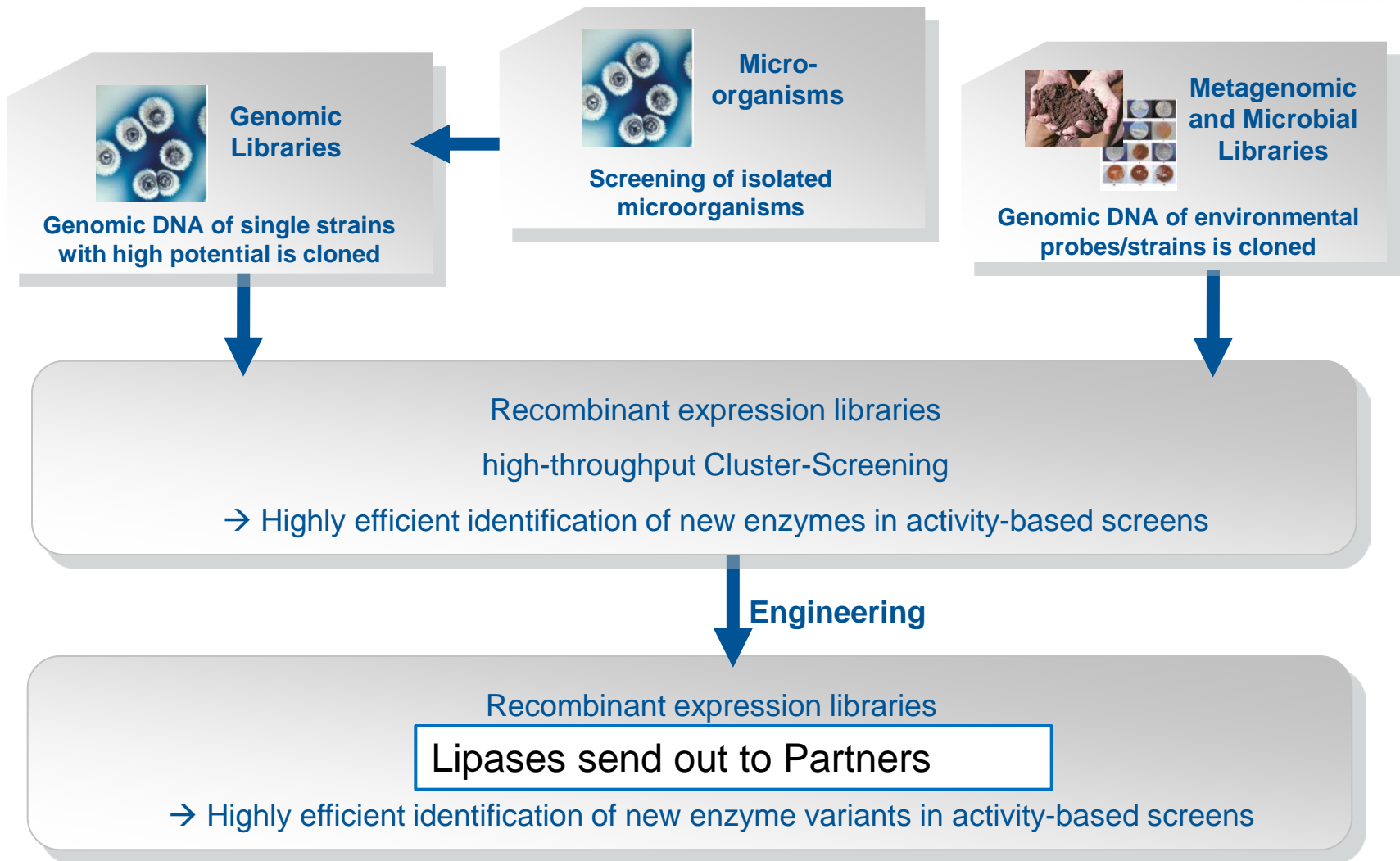




## WP3: Enzymatic synthesis and modification

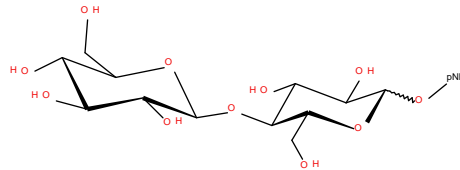
- enzymatic modification of natural biosurfactants to enhance surfactant performance
- enzymatic synthesis of novel types of glycolipid-biosurfactants derived from fatty acids, alkanols or terpenes in combination with rare sugars or oligosaccharides

# WP3: Screening for new enzymes

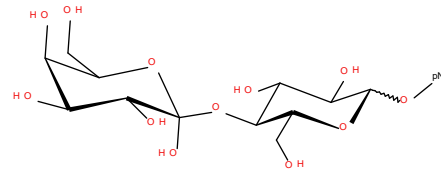


# WP3: Enzymatic glycolipid modification

- screening for transglycosidases
- established with pNP-derivatives of cellobiose and lactose and with pNP-derivatives of palmitic acid for lipases



pNP derivative of cellobiose

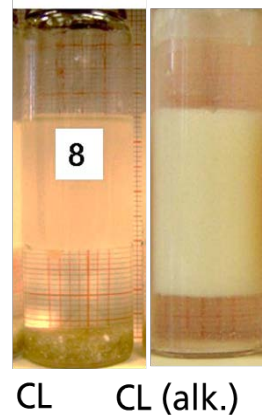
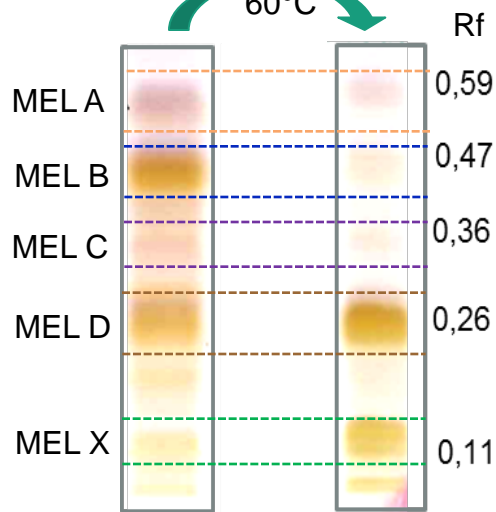
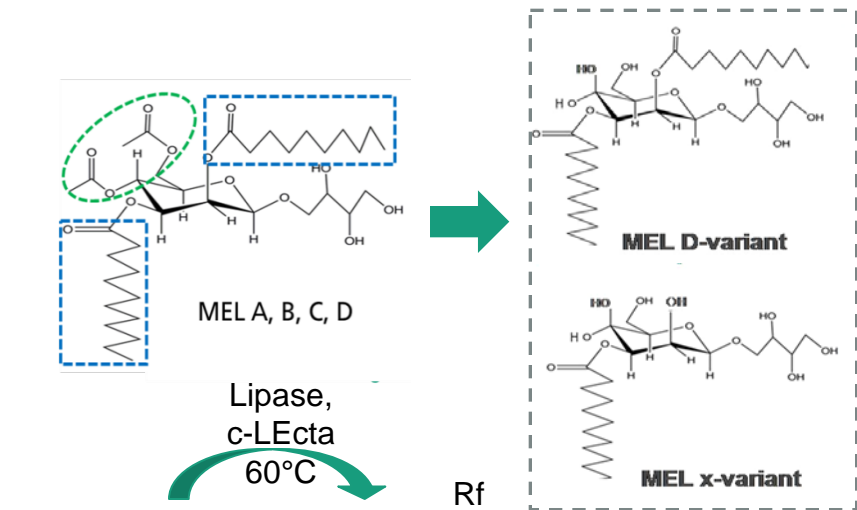


pNP derivative of lactose

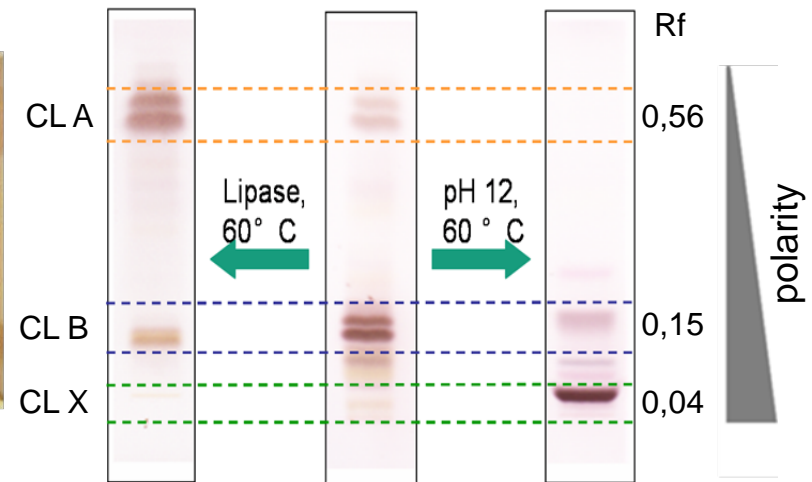
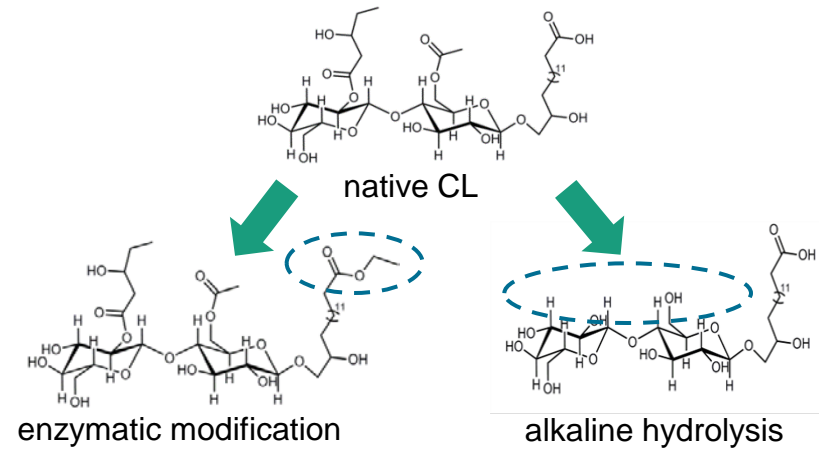
- screening of different genomic and metagenomic libraries (altogether 300,000 clones)
- four new glycosidases able to hydrolyze cellobiose- and lactose derivatives
- similarities to known glucosidases, glycosyl-hydrolase and rhamnosidases
- identities to known database enzymes between 48% and 96%
- enzymes are available

# WP3: Modification of glycolipids from fungi

## Modification of mannosylerythritol lipids

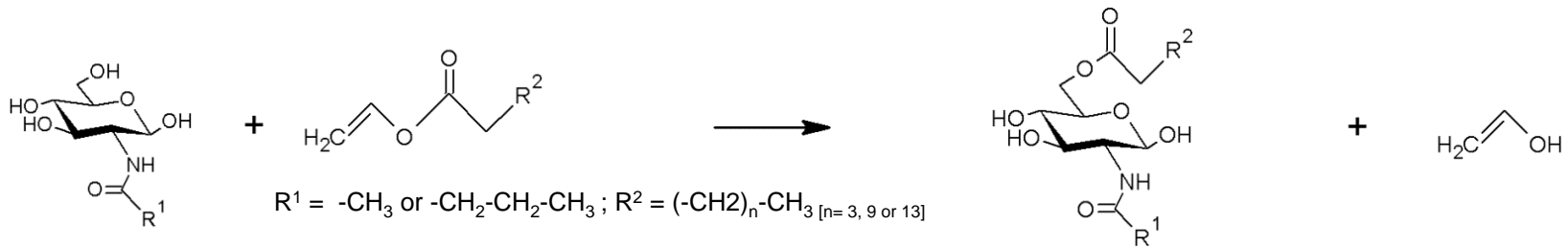


## Modification of cellobioselipids



# WP3: Enzymatic synthesis of novel glycolipids

## Different amino sugar fatty acid esters synthesized via lipase-catalyzed transesterification

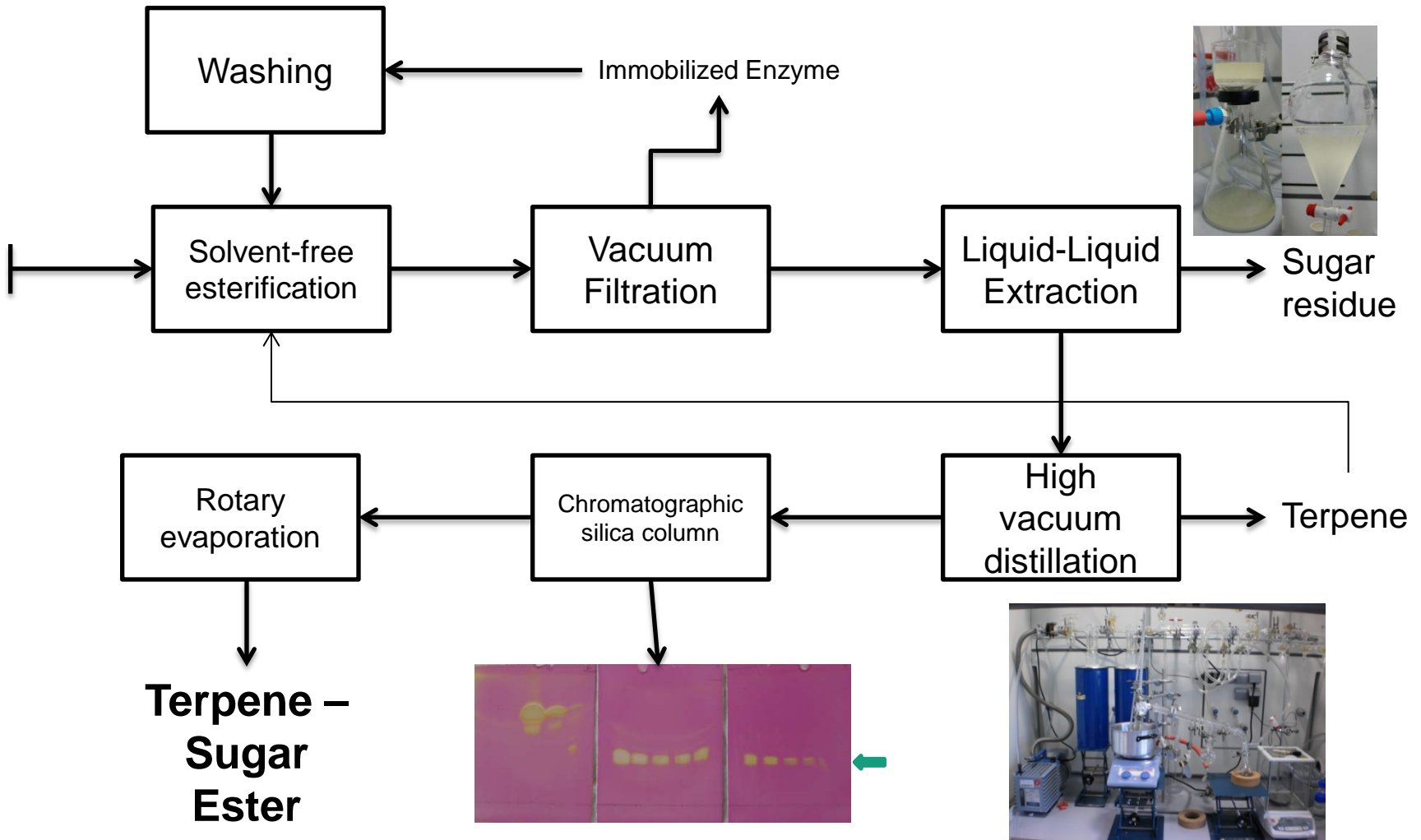


- Experiments carried out for 48h in 2m2b with Novozyme 435™ at 40°C and 300 rpm
- Structures elucidated via NMR
- Quantification using high performance liquid - and gas chromatography
- Purification via MPLC
- Influence on the synthesis of different parameters tested:
  - Substrate solubility
  - Water content
  - Substrate ratio
  - Temperature

# WP3: Enzymatic synthesis of novel terpenoid sugar esters

- Solvent-based and solvent-free process for novel, not yet characterized biosurfactants from terpenes and sugars
- Reaction kinetics determined
- Residual substrate and bio-catalyst recovery possible
- Analytical validation of products
- Promising surfactant properties of products
- Product stability at higher pH critical (hydrolysis)

# Solvent-free process



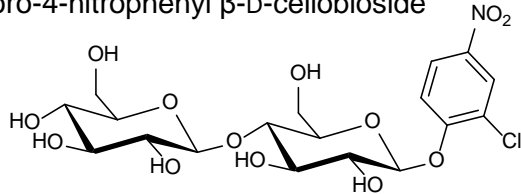
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Lars Wiemann

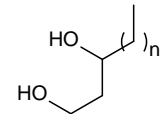
# WP3: Glycoceramidase-engineering for Synthesis



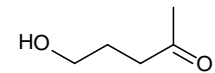
2-chloro-4-nitrophenyl β-D-cellobioside



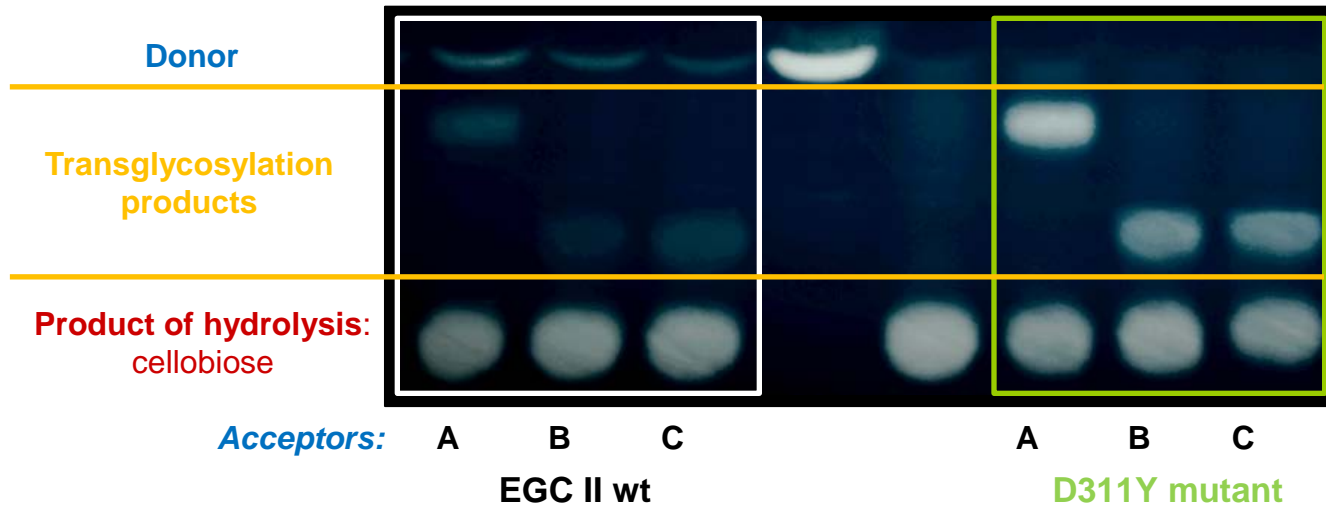
## Acceptors



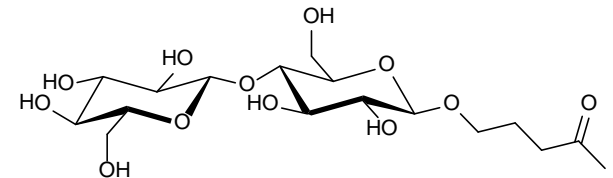
- (A) n = 4, octane-1,3-diol
- (C) n = 1, pentane-1,3-diol



- (B) 5-hydroxypentan-2-one



- (A and C): mixtures of regioisomers (62% for acceptor A)
- (B): **cellobiosidelipid 59%**



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Régis Fauré

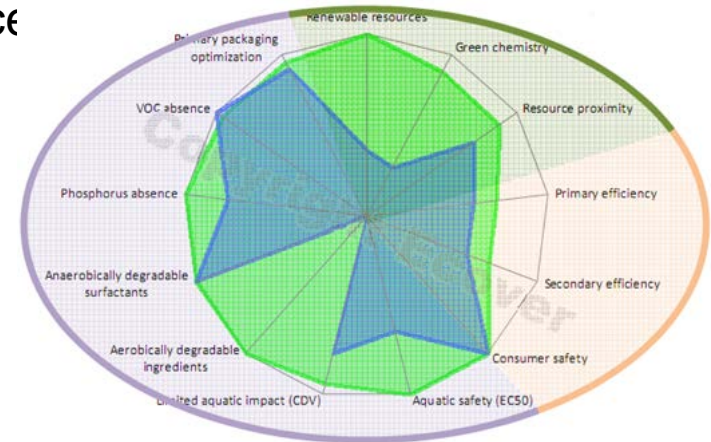
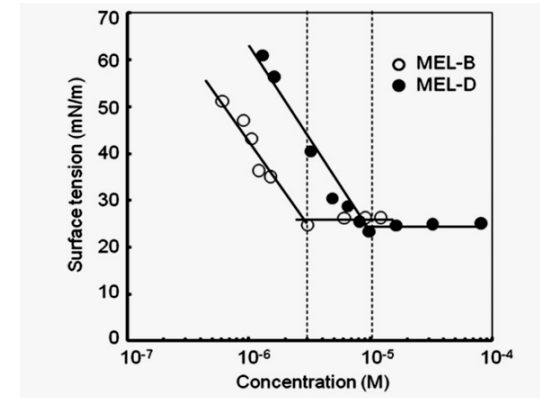


# WP4: Product characterisation and selection for product development

- Testing of the generated biosurfactants for basic physico-chemical properties
- Testing for antimicrobial properties (AS-HTS)
- Promising surfactants will be produced in 100 g scale for full application tests:

Foam quality, skin compatibility (HET-CAM-Test), dishwashing and laundry performance tests

Ecotoxicity and biodegradability of the new products



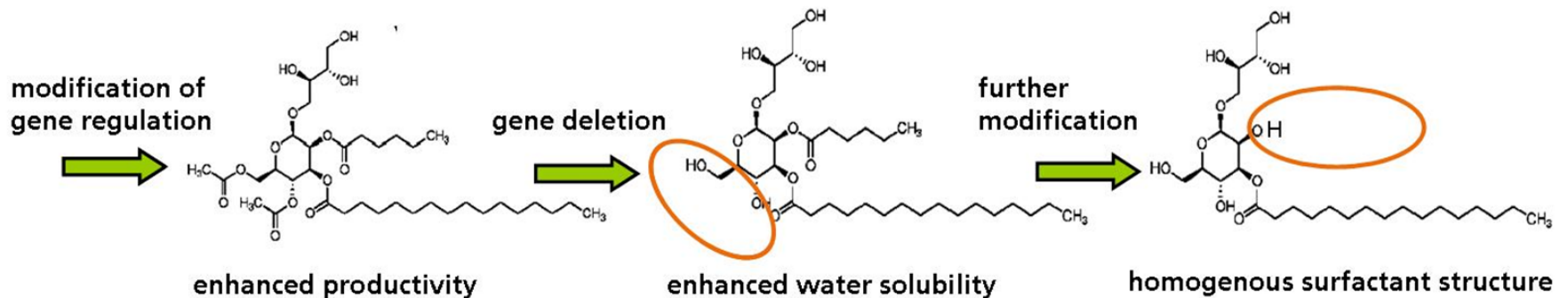
# WP4: Application tests performed

	CGE batch 1	GGE batch 1	MGE batch 1	CGE batch 2	MEL purified	CBL native	CBL hydrolysed
	Terpenol 1	Terpenol 2	Terpenol 3	Terpenol 4	Mannosylerythritol lipid	Cellulose lipid	Cellulose lipid
<b>Solubility in water</b>	pH adjustments needed	Water soluble, but unstable after one night at 4°C	Neutral pH	10000ppm	pH adjusted (pH=11), 5 days at 40°C	pH adjusted, 5 days at 60°C	Addition of hydrotrope (influences succeeding measurements)
<b>Min. surface tension measured [mN/m]</b>	27	26	31	37	29	40	
<b>Min. contact angle measured [°]</b>	49	30	59	58	27		
<b>CMC [ppm = mg/kg]</b>	2000	2000	>10000	>10000	100	1500	
<b>Foam</b>					No foam	No foam	
<b>Remarks</b>					Slow		

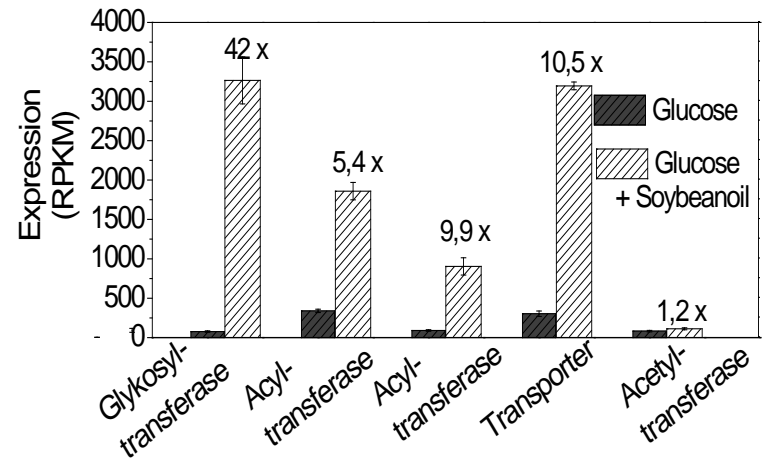
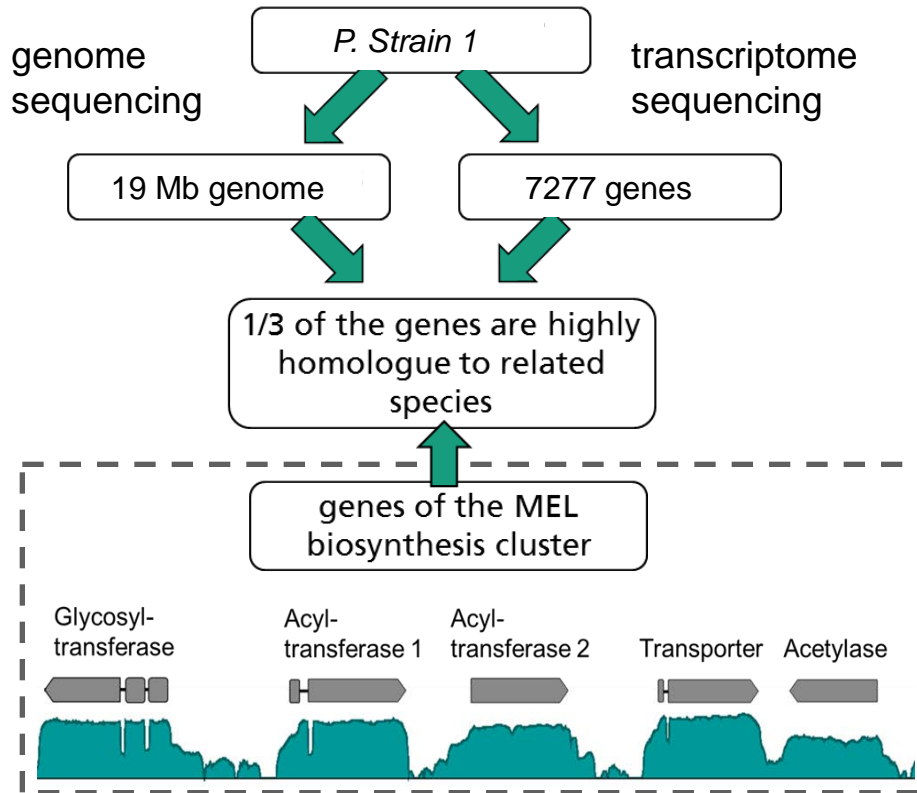
	Partial hydrolysed SL	Complete hydrolysed SL
<b>Solubility in water</b>	Yes	Yes
<b>Surface tension at CMC</b>	38 mN/m	31 mN/m
<b>Contact angle at CMC</b>	39°	35°
<b>Foam</b>	Foam, but low capability and stability	No foam with soft or hard water

# WP5: Metabolic engineering: Improved strains and surfactants for industrial application

- Identification of biosurfactant pathways
- Identification of regulatory mechanisms
- Engineering of biosynthetic pathways to obtain higher production yields and tailor surfactant structures



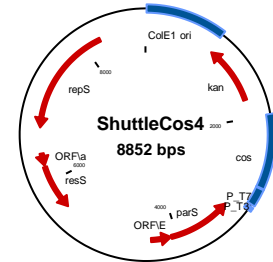
# WP5: Investigation of the MEL-metabolism in *P. strain1* on genome and transcriptome level



Differential expression analysis with and without a hydrophobic c-source reveals 13 upregulated metabolic groups including the MEL pathway

# WP5: Novel biosurfactant operons: Large insert libraries

- building of of a new shuttle cosmid
  - screenings in *E. coli* and *Bacillus* sp. possible
- protocols for high efficient transformation of cosmid libraries in screening host *B. subtilis* (surfactin-deficient) established
- Control library from *B. subtilis* (surfactin producer) built
- surfactin production of *B. subtilis* DSM was confirmed by emulsification tests
- screening was performed in DWP with MTP assay, Anthracene solubility assay and Methylene blue assay
- no biosurfactant positive hits were found



# WP6: Bioprocess Engineering and DSP

- Scale Up/Batchwise DSP
- Membrane design for In Situ Product Recovery and set up of an integrated process



# WP6: Downstream processing of MEL and CL

Biosurfactant	Purity	Consistency
Mannosylerythritol Lipid (MEL)	60-80 %	Viscous Fluid
Cellobiose Lipid (CL)	60-80 %	Powder



- Recovery and concentration of the samples
- Obtain a purity >> 90 %
  
- Procedures
  - Precipitation of the biosurfactants  
(→ not suitable, poor quality of the product)
  - Extraction with Supercritical CO<sub>2</sub>
  - Membrane filtration  
variation of membranes, solvents and ratios of the mixture  
biosurfactant / solvent

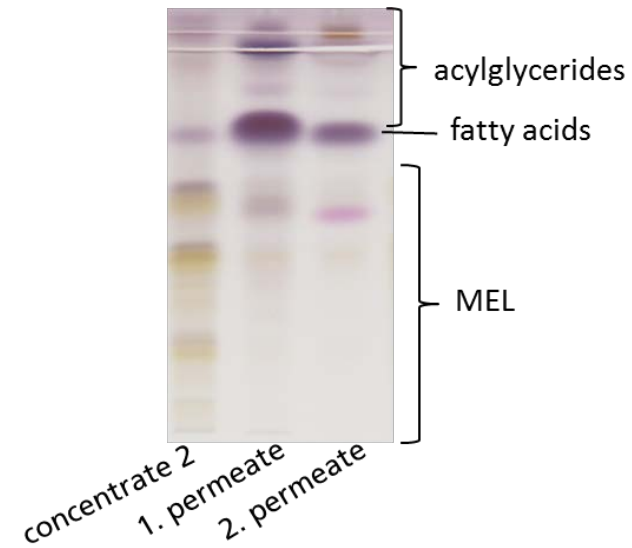
# DSP of MEL and CL: Membrane filtration

- Membrane filtration of MEL (63 % → 90 % purity)

- Suitable membrane, solvent and sample mixture for MEL identified
- two step filtration leads to 90 % purity

- Membrane filtration of CL (46 % → 90-100 % purity)

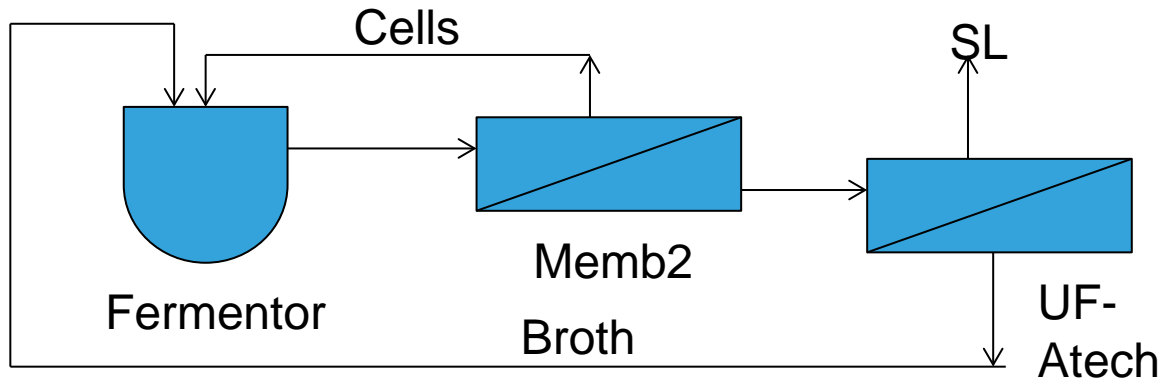
- highly selective separation



	Concentrate	Permeate
Membrane 1 + Ethylacetate		
% CL	87	0
% other lipids	13	100
Membrane 1 + Hexane		
% CL	100	0
% other lipids	0	100



# WP6: SL yield after membrane purification (ISPR)



	sediment	soluble	sediment + soluble
Membrane1	6%	63%	53%
Membrane2	30%	98%	78%



	sediment	soluble	sediment + soluble
Membrane1	2%	45%	41%
Membrane2	15%	91%	82%

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Bert Lemmens

# WP6: DSP on sophorolipids

In Situ Product Recovery on site:

- Installation of lab stack device at pilot facilities Ecover
- First trial a few months ago
- results limited due to technical difficulties
- Second trial scheduled for end of this month



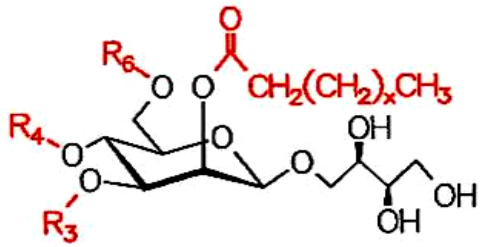
# WP7: Dissemination and Exploitation

- Presentations at workshops/trade fairs
  - Forum Industrial Biotechnology @ Biotechnika (11/2011)
  - Hannover Messe (4/2012) Exhibition @ Fraunhofer House of Sustainability
  - “Grüne Woche” Berlin 1/2013 and 1/2014, Exhibition @ Fraunhofer Booth
- First International Workshop on Biosurfactants  
16.-17.5.2013 @ DECHEMA, Frankfurt

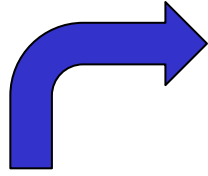
organized by BioSurf: Christoph Syldatk, Rudolph Hausmann, Steffen Rupp  
(see <http://events.dechema.de/biosurf.html>)

# Summary

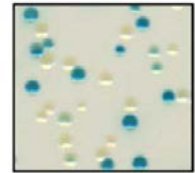
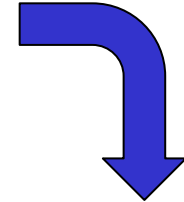
- Production of several modified bio-surfactants (SL, MEL, CL)
- Novel bio-surfactants identified and produced (Trehalose-Lipids, MEL-X)
- Novel bio-surfactants synthesized enzymatically (Terpenols, aminoglycosid lipids, cellobiose lipids)
- Identification of biosynthetic pathways for MEL production and their regulation using NGS
- Application tests and qualification of several novel bio-surfactants and their modifications (still ongoing)
- Novel DSP-Processes for SL, MEL and CL (still ongoing)
- Cost reduction in SL-production by ISPR-process



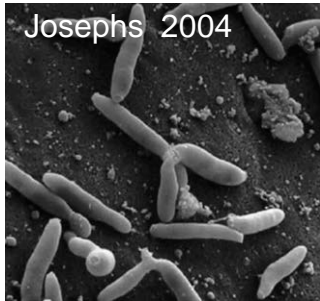
Identification and preparation



Bioprocess Engineering and DSP



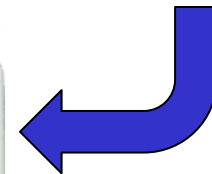
Selection of mutants by digital imaging screening



Thank you for your attention



product development



Dissemination and Exploitation

