

BIOSYNTHESIS OF TACROLIMUS

<u>Tacrolimus</u> is formed by three major PKS's and a NRPS followed by several modification enzymes



ERA-IB2 PROYECT INMUNOTEC

ROBUST FERMENTATION PROCESS FOR THE PRODUCTION OF TACROLIMUS

PARTNERS

1. Coordinators: Dr. Paloma Liras and Dr. Juan F. Martín - INBIOTEC Institute of Biotechnology (León,Spain): WP1, WP4

2. Dr. Wolfgang Wohlleben - Eberhand-Karls-Universität Tübingen (Tübingen, Germany): WP2

3. Dr. Marta Vaz Mendes - IBMC-Instituto de Biología Molecular e Celular (Porto, Portugal): WP3

4. Dr. Lutz Heide - Eberhand-Karls-Universität Tübingen (Tübingen, Germany) : WP5

5. Dr. Tania Velasco - ANTIBIÓTICOS S.A. (León, Spain): WP6

WORK PACKAGEs 1 and 4

PARTNER 1: INSTITUTE OF BIOTECHNOLOGY. LEÓN, SPAIN

Personnel:

Principal Investigator: Prof. Paloma Liras Prof. Juan F. Martín Dr. Antonio Rodríguez-García Dr. Fernando Santos Beneit María Ordoñez, Ph.D. student **Collaborator** :Prof. Hrvoje Petkovic (Slovenia)

WP1: Analysis of the sequences of S. tsukubaensis genome. Characterization of the tacrolimus gene cluster and regulatory genes



WP1. THE S. tsukubaensis GENOME CHARACTERISTICS

One linear chromosome (7.62 Mbp) and two circular plasmids of 24.7 and 31.1 kbp

It contains: 6623 protein-encoding large ORFs (>0.8kb) 6 rRNA operons 68 tRNAs 52 Sigma factors About 20 secondary metabolites clusters

Using the genome sequence WP1 studied and provided genes to the other partners:

Genes for tacrolimus biosynthesis subcloned The *bul* cluster for butyrolactone biosynthesis and regulation Genes of the ethylmalonyl-CoA pathway The *pho* regulon. Genes controlled by phosphate. Pho boxes (to WP4) The nitrogen metabolism structural and regulatory genes (to WP2) Genes related to oxidative stress (to WP3) Clusters for other secondary metabolites (to WP5)





Tacrolimus byosinthesis is regulated by two positive regulatory elements in *Streptomyces tsukubaensis* In colaboration with Prof. H Petkovic



fkbR

fkbN

fkbB

hrdAgu



fkbD fkbM fkbN

fkbQ fkbR 4462 4463 4464



Increasing *fkbN* copy numbers increase 70% production

Goranovic et al., 2012

Butyrolactones initiate the production of secondary metabolites by binding butyrolactone receptor proteins



bulY

The six genes were separatedly deleted. The effect on tacrolimus production was: bulR1 is essential. Deletion results in 80% decrease in production. bulR2 is not required bulS1 or bulS2 : 35-70% decrease bulZ or bulY: 50% increase in tacrolimus production, probably by cross-regulation

Salehi-Najafabadi et al. 2014



WP4: Phosphate control of primary metabolism and tacrolimus biosynthesis. Phosphate-deregulated mutants

Attempts to disrupt the phoP gene: only single cross-over recombinants

- 1. The Redirect approach: phoP replacement by the aac(3)/V cassette in a cosmid
- 2. The cytosine deaminase (codA) system/5-fluorocytosine: phoP replacement construction
- 3. The cytosine deaminase system *plus* a thermosensitive replicon (pSG5) (in progress)



No *phoP*-deleted mutants could be obtained by either approach



Transcriptomic studies to analyze the

Interaction of phosphate and carbon sources exerting carbon catabolite regulation of tacrolimus production in *S. tsukubaensis*



Carbon catabolite regulation at low phosphate concentration

•MG (2.5 mM Pi)

•10⁷ spores/ml, 28°C 220rpm

•Carbon additions at 70h (3%):

Glucose, Glycerol, Maltose, Sucrose, •Fructose, Lactose, Manitol and Xylose

•Bioassay with Saccharomyces cerevisiae

• Tacrolimus cuantification by HPLC



THREE MAIN TYPES OF TRANSCRIPT PATTERN OF GROUPS OF GENES UNDER PHOSPHATE-LIMITING CONDITIONS



IMMUNOTEC – WP2:

Eberhard-Karls-University of Tübingen, Germany Institute of Microbiology and Infection Medicine Depart. Microbiology and Biotechnology

Prof. Wolfgang Wohlleben Dr. Agnieszka Bera PhD student Annika Kemeny PhD student Susann



Nitrogen regulation of primary metabolism and tacrolimus biosynthesis. Mutants altered in nitrogen metabolism.

Strategies to optimize tacrolimus production by the modification of nitrogen metabolism

- 1. Construction of strains able to use nitrate, a substrate that enhance antibiotic production
- 2. Optimize the production of pipecolic acid, a tacrolimus precursor
- 3. Optimize lysine biosynthesis, a precursor of pipecolic acid

Regulation of nitrogen metabolism in S. coelicolor and S. tsukubaensis



S. tsukubaensis is unable to grow on nitrate due to impairment of the NO₃/NO₂ assimilation

Nitrogen sources used by S. tsukubaensis:

- ammonium
- glutamine
- glutamate
- arginine
- asparagine
- ...but no nitrate



Nitrate/nitrite assimilation pathway



Griess-Ilosvay-Assay: Nitrate to nitrite

S. coelicolor: active pathway



S. tsukubaensis: inactive pathway



S. tsukubaensis is unable to reduce nitrate to nitrite and then nitrite to ammonium

Model for NnaR dependent control of nitrate/nitrite assimilatory genes in *Streptomyces coelicolor*



NnaR dependent expression of the nitrate/nitrite assimilation genes is:

- activated during general nitrogen limitation
- activated in the presence of nitrate
- repressed in the presence of high ammonia concentrations

Streptomyces tsukubaensis pGM1190



Streptomyces tsukubaensis pGM1190/nnaR

HETEROLOGOUS EXPRESSION OF nnaR





S. tsukubaensis: Growth in MG + 100 mM Nitrate



Heterologous expression of NnaR initiated an activation of the NO₃/NO₂ assimilation pathway

II. Strategies to optimize the pipecolic acid precursor



1. Overexpression of the *fkbL* or *fkbLfkbP* increased tacrolimus production by 45%.

2. Heterologous expression of *pip* genes from:

- Streptomyces pristiniaspiralis (pipA)
- Actinoplanes friuliensis (pip)

resulted in significant increase of the tacrolimus producton by over 60%.



Optimization of the lysine biosynthesis in *S. tsukubaensis*

Homologs of *lysC* and *dapA* in *S. tsukubaensis* identified

Approaches:

- *lysC*^{*} (STSU_3111): site-directed mutagenesis (Ser→Tyr) in *lysC*_β region to get a feedback inhibition resistant aspartate kinase.
- Overexpression of *lysC** (STSU_3111) and *dapA* (STSU_1603) resulted in slight increase of the tacrolimus production.



 Simultaneous overexpression of *pip* and *lysC*/dapA* in S. tsukubaensis – on going





WP3. MODULATION OF OXIDATIVE STRESS

Partner 3:

Marta V. Mendes - Pl Sílvia Pires - PhD student Rute Oliveira - Research fellow

PORTO (PORTUGAL)



WP3. REACTIVE OXYGEN SPECIES (ROS) AND DETOXIFICANT ENZYMES





WP3. Antioxidant defences in *S. tsukubaensis* (MGm medium)



Native-PAGE stained for catalase activity



Native-PAGE stained for SOD activity







Tacrolimus production by the mutants



- 1 Ascorbic acid (50mM)
- 2 Ascorbic acid (50mM) + H_2O_2 (1M)
- $3 H_2O_2$ (1M)
- 4 FK-506 (0,25μg/μL)
- 5 FK-506 (0,25μg/μL) + H₂O₂ (1M)



Expression by PCR in the *∆sodA* mutant of the tacrolimus genes oxidative stress genes Iron uptake metabolism Phosphate metabolism



[Saccharomyces cerevisae BY4741]

TAC production (% of WT)







OxyR redox activation (LysR-type)





H₂O₂ sensitivity

WT



3.47 cm ± 0.229

 $\Delta oxyR$



4.65 cm ± 0.069

∆ahpC



2% YED Medium 9 M H₂O₂



wt_vs_Δ*oxyR* : 47 differences

wt_vs_Δ*ahpC* : 66 differences



2.73 cm ± 0.068

ΔoxyR_vs_ΔahpC: 46 differences

Positive regulation: 22 genes, e.g aphC Negative regulation: 10 genes, e.g *oxyR*



The oxyR regulon







1-*sig-oxyR* promoter region

2-padR-ahpC promoter region

WP5. HETEROLOGUS EXPRESSION OF THE TACROLIMUS BIOSYNTHETIC GENE CLUSTER IN MODIFIED Streptomyces STRAINS

Eberhard Karls Universitat Tübingen

Overexpression of the tacrolimus cluster. Heterologous systems. Superhosts

Partner 4: Prof. Lutz Heide Dr. Adam Jones Dr. Bertolt Gust Dr. Christian Appel



I. Cloning of FK506 (tacrolimus) gene cluster into P1-derived phage artificial chromosome (PAC)





Cloning of FK506 (tacrolimus) gene cluster into phage artificial chromosome (PAC)



Bundesministerium für Bildung und Forschung





ERA

B



Bundesministerium für Bildung und Forschung





Optimization of tacrolimus production in *S. coelicolor*



Bundesministerium für Bildung und Forschung



Heterologous expression of the bafilomycin-like gene cluster from *S. tsukubaensis*

- A family of 16-membered ring macrolide antibiotics.
- Potent vacuolar H+-ATPase inhibitors
 - antifungal, immunosuppressant, antitumor and antiparasitic (Yu et al, 2011)
 - Employed to study ATPase

ERA

- Produced by various actinomycetes
 - S. griseus for bafilomycins A1, A2, B1, B2, C1 and C2
 - Kitasatospora setae for bafilomycin B1
 - Streptomyces lohii



Bafilomycin B2 (7 $R_4 = CH_3$)

Homologous gene clusters	
All hits	✓ Download graphic
Query sequence	
NC_016109.1_c27	Kitasatospora setae KM-6054, complete genome.
GU390405.1_c1	Streptomyces lohii strain ATCC BAA-1276 bafilomycin Ł

Bafilomycins

CONCLUSSIONS

S. tsukubaensis genome has been sequenced and information provided to all the partners

The *bul* DNA region for butyrolactone biosynthesis and receptors and their binding to specific promoter sequences has been analyzed

The *pho* DNA region has been analyzed and transcriptomic analysis on the CCR response in low phosphate concentration, favoruable for tacrolimus production, has been studied

The genes for nitrogen asimilation and their regulators have been analyzed. S. tsukubaensis transformants, carrying the S. coelicolor nnaR gene, able to grow on nitrate have been obtained

The genes involved in oxidative stress regulation have been studied and related to tacrolimus production

Tacrolimus overproducer strains have been obtained by transformation with the *fkbN* gene, the *pipA* gene o by knock out of the *aphC* gene. This results in 40 to 60% increase production in each case.

The tacrolimus gene cluster of *S. tsukubaensis* has been expressed in *S. coelicolor* and the heterologous production has been optimized 80% over the original production

PUBLICATIONS : 4 published or in press, others submitted

Martínez-Castro, M., Salehi-Najafabadi Z., Romero, F., Pérez-Sanchis, R., Fernández-Chimeno R.I., Martín, J.F., Barreiro, C. (2013). Taxonomy and chemically semi-defined media for the analysis of the tacrolimus producer *Streptomyces tsukubaensis*. Applied Microbiology and Biotechnology 97:2139-2152. **WP1**

Goranovič D, Blažič M, Magdevska V, Horvat J, Kuščer E, Polak T, Santos-Aberturas J, Martínez-Castro M, Barreiro C, Mrak P, Kopitar G, Kosec G, Fujs S, Martín JF, Petković H. (2012) FK506 biosynthesis is regulated by two positive reglatory elements in *Streptomyces tsukubaensis*. <u>BMC</u> Microbiol. 12:238. **WP1**

Jones AC, Gust B, Kulik A, Heide L, Buttner MJ, Bibb MJ (2013) Phage P1-derived artificial chromosomes facilitate heterologous expression of the FK506 gene cluster. PLoS One 8: e69319. **WP5.**

Salehi-Najafabadi, Z., Barreiro C, A. Rodríguez-García A, A. Cruz A, López GR, Martín JF (2014) The γ -butyrolactone receptors BulR1 and BulR2 of *Streptomyces tsukubaensis* control the butyrolactone synthetases and the production of tacrolimus: Characterization of BulR1 DNA-binding sequences. In press. **WP1**

Kocadinc S, Wohlleben W. and A. Bera. (2014) Optimization of the N-containing precursor supply by genetic engineering of *Streptomyces tsukubaensis* for FK506 production improvement. Manuscript in preparation. **WP2**

Pires S, R. Oliveira, T. Beites, P. Moradas-Ferreira and M.V. Mendes (2014) The OxyR-dependent regulatory mechanisms of oxidative stress response and iron metabolism interplaywith tacrolimus production in *Streptomyces tsukubaensis*. Send for publication. **WP3**

Jones A. C., Flinspach K., Herbig A., Apel A. K., Nieselt K. and Heide L. (2014) RNA-seq transcriptional analysis of the FK506 biosynthetic gene cluster in *Streptomyces tsukubaensis* NRRL118488. International Microbiology. Submited, under revisión. **WP5**

Heterologous expression of S. coelicolor nnaR in S. tsukubaensis

Expression of nitrate/nitrite assimilatory genes is:

- activated during general nitrogen limitation
- activated in the presence of nitrate
- repressed in the presence of high ammonium conc.



Heterologous expression of nnaR restored nitrate assimilation in S. tsukubaensis



S. tsukubaensis growth on MG suppl. with 100mM nitrate

Streptomyces tsukubaensis pGM1190



Heterologous expression of NnaR initiated an activation of the NO₃/NO₂ assimilation pathway